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Dynamic properties of motor proteins with two subunits

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Abstract

The dynamics of motor protein molecules consisting of two subunits is investigated using simple discrete stochastic models. Exact steady-state analytical expressions are obtained for velocities and dispersions for any number of intermediate states and conformations between the corresponding binding states of proteins. These models enable us to provide a detailed description and comparison of two different mechanisms of the motion of motor proteins along the linear tracks: the hand-over-hand mechanism, when the motion of subunits alternate; and the inchworm mechanism, when one subunit is always trailing another one. It is shown that the proteins in the hand-over-hand mechanism. The effect of external forces on dynamic properties of motor proteins is also discussed. Finally, a quantitative method, based on experimental observations for single motor proteins, is proposed for distinguishing between two mechanisms of motion.

1. Introduction

Motor proteins, also called molecular motors, are active enzyme molecules that play a fundamental role in most biological processes, but especially in cellular transport, motility, cell division, and transcription [1–3]. Motor proteins, such as kinesins, dyneins, myosins, polymerases, helicases, etc, function by stepping between equally spaced binding sites along the rigid polar linear tracks (microtubules, actin filaments, DNA molecules), and the motion is powered by the hydrolysis of adenosine triphosphate (ATP) or related compounds. The mechanisms of the transformation of chemical energy of hydrolysis into the mechanical work in motor proteins are not yet fully understood [3].

Recent experimental advances have allowed for the determination of structural and dynamic properties of motor proteins with a high degree of precision at a single-molecule

level [4–16]. Crystal structures suggest that many motor proteins have two domains elastically coupled together, each capable of hydrolysing ATP and moving along the linear track [3, 5]. Two possible mechanisms of the coordinated motion of protein molecules with two motor heads have been proposed [3]. In a hand-over-hand mechanism, at each step only one motor head undergoes a sequence of mechanochemical transitions so that the motor subunits alternate between trailing and leading positions at the beginning of the cycle. In this mechanism the motor subunits are fully equivalent to each other. In contrast, according to an inchworm mechanism, one motor domain is always ahead of the other one during the cycle; i.e., the motor heads are not equivalent at all times. Experiments on single-molecule fluorescently labelled myosins V, that step along the actin filaments, and on kinesins, that move along microtubules, support the hand-over-hand mechanism for these motor proteins [12, 13, 15, 16]. However, there are indications that dyneins probably utilize the inchworm mechanism [17].

Successes in experimental studies have strongly stimulated many theoretical investigations of mechanisms and dynamics of molecular motors [18-31]. Most theoretical work on motor proteins follows two main directions. One approach utilizes the concept of thermal ratchets [19, 28, 30]. According to this idea, the motor protein is viewed as a Brownian particle that moves in two different periodic but asymmetric potentials, switching stochastically between them. This method takes into account the internal structure and interactions between different domains in protein molecules; however, the results are mainly numerical and depend on the specific potentials used in calculations. An alternative approach is based on multi-state discrete stochastic (chemical kinetic) models [18, 20-26, 29, 31]. In this method, it is assumed that the motor proteins move sequentially between different molecular conformations and states. The molecular motors are associated with particles that move along one-dimensional periodic lattices with different forward and backward rates. The lattices correspond to biochemical pathways for the motor proteins, while the sites in the period describe the biochemical cycle when the protein molecule travels between two consecutive binding sites. Using this mapping of the motion of a random walker and applying the method of Derrida [32], exact analytical expressions for the mean velocities and dispersions are derived for any number of intermediate states (i.e., for the period of any size) and for different complexity of biochemical pathways [21–25]. It was demonstrated that this approach allowed for the successful analysis of the full dynamics of single kinesin and myosin V molecules [26, 29]. However, the weakness of this method is the fact that the internal structure of motor proteins, namely, the motion and interactions of different subunits, is not taken into account.

Determining how the different motor heads move relative to each other is critical for the overall understanding of a motor protein's dynamics and functions. Current experimental methods with single-molecule fluorescent labels, that distinguish between the different types of molecular motion, require a detailed knowledge of the protein structure, which is not always available. In addition, the labelled proteins may change their biochemical properties in comparison with the original species. However, it would be more advantageous to use simpler less-invasive experimental methods along with better theoretical models to study the specific mechanisms of molecular motors. In this article we investigate the dynamics of motor proteins by developing a set of simple multi-state discrete stochastic models. In our approach the motor proteins consist of two interacting particles that correspond to different motor subunits in real enzymes. Explicit formulae for the velocities and dispersions are obtained for two different mechanisms of motion. We suggest a method to distinguish between two possible mechanisms by analysing time trajectories of single motor proteins obtained in optical-trap experiments [6–11] in combination with the bulk biochemical kinetic data.



Figure 1. General schematic view of periodic stochastic models of motor proteins consisting of two subunits. Two parts of the molecular motor cannot occupy the same site and cannot be more than *m* sites apart. The motor domain at site *j* can make a forward or backward step with the rate u_j or w_j , correspondingly, if these transitions are allowed by another motor domain.

2. Theoretical approach

2.1. General model

Consider a motor protein molecule with two subunits that travels along the filament track. We model this system as two identical interacting particles moving on a periodic one-dimensional lattice, as shown in figure 1. There are N intermediate discrete states on a biochemical pathway between two consecutive binding sites. In the simplest approximation, we assume that the particles interact through hard-core exclusion, i.e., they cannot occupy the same site. Also, the particles cannot run away from each other. If x_1 and x_2 are the positions of the motor particles then

$$|x_1 - x_2| \leqslant m,\tag{1}$$

where *m* is an integer that specifies how far apart two motor domains can be found in the protein molecule. In the lattice the sites $x = \pm Nl$ (l = 0, 1, ...) correspond to binding sites of the molecular motor. The distance between two consecutive binding sites is *d*, which is equal to 8.2 nm for kinesins and dyneins moving on microtubules and 36 nm for myosins V and VI travelling along the actin filaments [1–3].

The particle at site *j* moves forward (backward) with the rate u_j (w_j) if the site j + 1 (j - 1) is available and the move does not violate the condition (1): see figure 1. Because of periodicity the transition rates are related, $u_{j\pm Nl} = u_j$ and $w_{j\pm Nl} = w_j$ for l = 0, 1... and $0 \le j \le N - 1$. The dynamic properties of motor proteins are specified by the drift velocity

$$V = V(\{u_j, w_j\}) = \lim_{t \to \infty} \frac{\mathrm{d}}{\mathrm{d}t} \langle x(t) \rangle, \tag{2}$$

and dispersion (or diffusion constant)

$$D = D(\{u_j, w_j\}) = \lim_{t \to \infty} \frac{1}{2} \frac{\mathrm{d}}{\mathrm{d}t} [\langle x^2(t) \rangle - \langle x(t) \rangle^2], \tag{3}$$

where x(t) is the position of the centre of mass of the protein molecule at time t. It is convenient to express the degree of fluctuations of the molecular motor in terms of a dimensionless function called randomness [4]

$$r = \frac{2D}{\mathrm{d}V}.\tag{4}$$

This function sets bounds on the number of rate-limiting biochemical transitions and thus yields important information about the mechanism of a motor protein's processivity [8, 23, 26].



Figure 2. The general picture of the hand-over-hand mechanism for the motor proteins with two heads. Each motor moves along its own discrete lattice. While the trailing grey head jumps, the black leading head does not move. The distance between the neighbouring binding sites for each subunit is 2*d*. Arrows indicate the allowed transitions.

The motor proteins in experiments and in the cellular environment frequently work against external loads [1-3]. External forces modify the transition rates in the following way [21-25]:

$$u_j(F) = u_j(0) \exp\left(-\frac{\theta_j^+ F d}{k_{\rm B}T}\right), \qquad w_j(F) = w_j(0) \exp\left(+\frac{\theta_j^- F d}{k_{\rm B}T}\right), \tag{5}$$

where $\sum_{j=0}^{N-1} (\theta_j^+ + \theta_j^-) = 1$, and θ_j^{\pm} are load-distribution factors that specify how the external load changes the energy activation barriers for the biochemical transitions from the state *j*.

The dynamic properties of the motor proteins with two domains depend on the specific mechanism of the motion. Below we consider in detail the hand-over-hand and the inchworm mechanisms.

2.2. Hand-over-hand mechanism

In this mechanism, the trailing subunit makes N intermediate steps and becomes the leading particle, as shown in figure 2. Then the next particle makes N transitions. During the cycle each head advances the distance 2d so the centre of mass of the protein molecule moves only the distance d. This mechanism then can be viewed as a motion of two particles on periodic parallel one-dimensional lattices, where the distance between neighbouring binding sites is 2d. Because the particles are identical, this picture is easily mapped into the motion of the single particle (centre of mass) on the original one-dimensional lattice, for which the dynamics is well understood [32, 21].

At large times the exact expression for the drift velocity is given by [32, 21]

$$V_{\rm hoh} = d \left(1 - \prod_{j=0}^{N-1} \frac{w_j}{u_j} \right) / R_N, \tag{6}$$

and

$$R_N = \sum_{j=0}^{N-1} r_j, \qquad r_j = u_j^{-1} \left[1 + \sum_{k=1}^{N-1} \prod_{j+1}^{j+k} \frac{w_i}{u_i} \right].$$
(7)

The corresponding expression for dispersion can be written as [32, 21]

$$D_{\rm hoh} = \frac{d}{N} \left(\frac{dU_N + VS_N}{R_N^2} - \frac{(N+2)V}{2} \right), \tag{8}$$



Figure 3. Two possible configurations for the inchworm mechanism of the motion with m = 2. The black particle is always leading. The allowed transitions are indicated by arrows. The parameter *l* labels different binding sites.

where the auxiliary functions are

$$S_N = \sum_{j=0}^{N-1} s_j \sum_{k=0}^{N-1} (k+1) r_{k+j+1}, \qquad U_N = \sum_{j=0}^{N-1} u_j r_j s_j, \tag{9}$$

and

$$s_j = u_j^{-1} \left[1 + \sum_{k=1}^{N-1} \prod_{j=1}^{j-k} \frac{w_{i+1}}{u_i} \right].$$
(10)

It can also be demonstrated that in this mechanism $r_{\text{hoh}} > 1/N$ for any set of transitions rates $\{u_j, w_j\}$ [8, 23, 26].

2.3. Inchworm mechanism

Now consider the inchworm mechanism of the motion of the motor proteins with two subunits. In this mechanism one particle is always leading the other one, as shown in figure 3. In the simplest approximation, we discuss only the case m = 2 in equation (1), i.e., the two motor particles can only be found on two nearest-neighbour sites or next-nearest-neighbour sites: see figure 3.

To determine the dynamic properties of motor proteins in this model we develop a method that generalizes the original Derrida's approach [32, 23, 24]. The first step is to introduce $P_{j,k}(l, t)$ which is the probability of finding the trailing subunit of the motor protein molecule in state j and the leading subunit in state k (k = j + 1 or j + 2) with the trailing head at site l at time t (see figure 3). Here the parameter l labels the motor binding sites. Between two binding sites there are N intermediate protein states, labelled by $j = 0, 1, \ldots, N - 1$. The time evolution of this probability is governed by Master equations for $1 \le j < N - 1$:

$$\frac{\mathrm{d}P_{j,j+1}(l,t)}{\mathrm{d}t} = u_{j-1}P_{j-1,j+1}(l,t) + w_{j+2}P_{j,j+2}(l,t) - (u_{j+1}+w_j)P_{j,j+1}(l,t)$$

$$\frac{\mathrm{d}P_{j,j+2}(l,t)}{\mathrm{d}t} = u_{j+1}P_{j,j+1}(l,t) + w_{j+1}P_{j+1,j+2}(l,t) - (u_j+w_{j+2})P_{j,j+2}(l,t).$$
(11)

The Master equations are slightly different for j = 0,

$$\frac{\mathrm{d}P_{0,1}(l,t)}{\mathrm{d}t} = u_{N-1}P_{N-1,1}(l-1,t) + w_2P_{0,2}(l,t) - (u_1+w_0)P_{0,1}(l,t)$$

$$\frac{\mathrm{d}P_{0,2}(l,t)}{\mathrm{d}t} = u_1P_{0,1}(l,t) + w_1P_{1,2}(l,t) - (u_0+w_2)P_{0,2}(l,t).$$
(12)

Similarly, for j = N - 1 we have

$$\frac{dP_{N-1,0}(l-1,t)}{dt} = u_{N-2}P_{N-2,0}(l-1,t) + w_1P_{N-1,1}(l-1,t) - (u_0 + w_{N-1})P_{N-1,0}(l-1,t) \frac{dP_{N-1,1}(l-1,t)}{dt} = u_0P_{N-1,0}(l-1,t) + w_0P_{0,1}(l,t) - (u_{N-1} + w_1)P_{N-1,1}(l-1,t).$$
(13)

Because of the conservation of probability, we require

$$\sum_{l=-\infty}^{+\infty} \sum_{j=0}^{N-1} [P_{j,j+1}(t) + P_{j,j+2}(l,t)] = 1 \quad \text{for all } t.$$
 (14)

Next, following Derrida's method [32], we define auxiliary functions

$$B_{j,j+1}(t) \equiv \sum_{l=-\infty}^{+\infty} P_{j,j+1}(l,t), \qquad B_{j,j+2}(t) \equiv \sum_{l=-\infty}^{+\infty} P_{j,j+2}(l,t),$$
(15)

and

$$C_{j,j+1}(t) \equiv \sum_{l=-\infty}^{+\infty} (j+Nl) P_{j,j+1}(l,t),$$

$$C_{j,j+2}(t) \equiv \sum_{l=-\infty}^{+\infty} (j+Nl) P_{j,j+2}(l,t).$$
(16)

Using the Master equations (11) we derive

$$\frac{\mathrm{d}}{\mathrm{d}t}B_{j,j+1}(t) = u_{j-1}B_{j-1,j+1}(t) + w_{j+2}B_{j,j+2}(t) - (u_{j+1} + w_j)B_{j,j+1}(t),$$

$$\frac{\mathrm{d}}{\mathrm{d}t}B_{j,j+2}(t) = u_{j+1}B_{j,j+1}(t) + w_{j+1}B_{j+1,j+2}(t) - (u_j + w_{j+2})B_{j,j+2}(t);$$
(17)

and

$$\frac{d}{dt}C_{j,j+1}(t) = u_{j-1}C_{j-1,j+1}(t) + w_{j+2}C_{j,j+2}(t) - (u_{j+1} + w_j)C_{j,j+1}(t)
+ u_{j-1}B_{j-1,j+1}(t)$$
(18)
$$\frac{d}{dt}C_{j,j+2}(t) = u_{j+1}C_{j,j+1}(t) + w_{j+1}C_{j+1,j+2}(t) - (u_j + w_{j+2})C_{j,j+2}(t)
+ w_{j+1}B_{j+1,j+2}(t).$$

In the limit of $t \to \infty$, again following Derrida's suggestions [32], we introduce the ansatz

$$B_{j,k}(t) \to b_{j,k}, \qquad C_{j,k}(t) - a_{j,k}t \to T_{j,k}.$$
(19)

Note that the parameters $b_{j,k}$, $a_{j,k}$ and $T_{j,k}$ are periodic, i.e., $b_{j,k} = b_{j+N,k+N}$, $a_{j,k} = a_{j+N,k+N}$ and $T_{j,k} = T_{j+N,k+N}$. Now define two new functions,

$$f_{j-1}^{(1)} \equiv w_j b_{j,j+1} - u_{j-1} b_{j-1,j+1}, \qquad f_{j+1}^{(2)} \equiv w_{j+2} b_{j,j+2} - u_{j+1} b_{j,j+1}.$$
(20)

At steady state $\frac{dB_{j,k}(t)}{dt} = 0$, and equations (17) transform into

$$0 = u_{j-1}b_{j-1,j+1} + w_{j+2}b_{j,j+2} - (u_{j+1} + w_j)b_{j,j+1},$$

$$0 = u_{j+1}b_{j,j+1} + w_{j+1}b_{j+1,j+2} - (u_j + w_{j+2})b_{j,j+2}.$$
(21)

Substituting (20) into these equations, we obtain

$$f_{j}^{(1)} = w_{j+1}b_{j+1,j+2} - u_{j}b_{j,j+2} = f_{0},$$

$$f_{j+1}^{(2)} = w_{j+2}b_{j,j+2} - u_{j+1}b_{j,j+1} = f_{0},$$
(22)

where f_0 is a constant. Then it can be shown that $f_j^{(1)} = f_{j-1}^{(1)} = f_0 = f_j^{(2)} = f_{j+1}^{(2)}$. This leads to the following expression for $b_{j,k}$:

$$b_{j,j+1} = \frac{-f_0}{u_{j+1}} + \frac{w_{j+2}}{u_{j+1}} b_{j,j+2} = \frac{-f_0}{u_{j+1}} \left[1 + \frac{w_{j+2}}{u_j} \right] + \frac{w_{j+1}w_{j+2}}{u_j u_{j+1}} b_{j+1,j+2},$$

$$b_{j,j+2} = \frac{-f_0}{u_j} + \frac{w_{j+1}}{u_j} b_{j+1,j+2} = \frac{-f_0}{u_j} \left[1 + \frac{w_{j+1}}{u_{j+2}} \right] + \frac{w_{j+1}w_{j+3}}{u_j u_{j+2}} b_{j+1,j+3}.$$
(23)

Solving these equations by iteration, and using the periodicity and the normalization condition,

$$\sum_{j=0}^{N-1} (b_{j,j+1} + b_{j,j+2}) = 1,$$
(24)

we finally derive

$$b_{j,k} = \frac{r_{j,k}}{R_N}, \qquad R_N = \sum_{j=0}^{N-1} [r_{j,j+1} + r_{j,j+2}],$$
 (25)

where

$$r_{j,j+1} = \frac{1}{u_{j+1}} \left\{ 1 + \sum_{k=1}^{N-1} \prod_{i=j}^{j+k-1} \left(\frac{w_{i+1}w_{i+2}}{u_i u_{i+2}} \right) + \frac{w_{j+2}}{u_j} \left[1 + \sum_{k=1}^{N-1} \prod_{i=j}^{j+k-1} \left(\frac{w_{i+1}w_{i+3}}{u_{i+1} u_{i+2}} \right) \right] \right\},$$

$$r_{j,j+2} = \frac{1}{u_j} \left\{ 1 + \sum_{k=1}^{N-1} \prod_{i=j}^{j+k-1} \left(\frac{w_{i+1}w_{i+3}}{u_{i+1} u_{i+2}} \right) + \frac{w_{j+1}}{u_{j+2}} \left[1 + \sum_{k=1}^{N-1} \prod_{i=j}^{j+k-1} \left(\frac{w_{i+2}w_{i+3}}{u_{i+1} u_{i+3}} \right) \right] \right\}.$$
(26)

To determine the coefficients $a_{j,k}$ and $T_{j,k}$, the ansatz (19) is substituted into equations (18) in the limit of large times. This yields the following equations:

$$0 = u_{j-1}a_{j-1,j+1} + w_{j+2}a_{j,j+2} - (u_{j+1} + w_j)a_{j,j+1},$$

$$0 = u_{j+1}a_{j,j+1} + w_{j+1}a_{j+1,j+2} - (u_j + w_{j+2})a_{j,j+2}.$$
(27)

Also the parameters $T_{j,k}$ must satisfy

$$a_{j,j+1} = u_{j-1}T_{j-1,j+1} + w_{j+2}T_{j,j+2} - (u_{j+1} + w_j)T_{j,j+1} - u_{j-1}b_{j-1,j+1},$$

$$a_{j,j+2} = u_{j+1}T_{j,j+1} + w_{j+1}T_{j+1,j+2} - (u_j + w_{j+2})T_{j,j+2} - w_{j+1}b_{j+1,j+2}.$$
(28)

Comparing equations (27) with equations (21) we conclude that $a_{j,k} = Ab_{j,k}$. The coefficient *A* can be found using the normalization condition (24) and it is equal to

$$A = \sum_{j=0}^{N-1} \frac{u_j r_{j,j+2} - w_j r_{j,j+1}}{R_N} = N \frac{1 - \left(\prod_{j=0}^{N-1} \frac{w_j}{u_j}\right)^2}{R_N}.$$
 (29)

To calculate the coefficients $T_{j,k}$ we introduce another set of auxiliary functions

$$y_{j+1}^{(1)} \equiv w_{j+2}T_{j,j+2} - u_{j+1}T_{j,j+1}, \qquad y_{j-1}^{(2)} \equiv w_jT_{j,j+1} - u_{j-1}T_{j-1,j+1}.$$
(30)

Then equations (28) can be rewritten in the following form:

$$a_{j,j+1} = y_{j+1}^{(1)} - y_{j-1}^{(2)} + u_{j-1}b_{j-1,j+1},$$

$$a_{j,j+2} = y_{j}^{(2)} - y_{j+1}^{(1)} - w_{j+1}b_{j+1,j+2}.$$
(31)

As shown in [23, 24], these equations are solved to yield the functions $y_i^{(1)}$ and $y_i^{(2)}$

$$y_{j}^{(1)} = -a_{j-1,j+1} + \frac{A}{N} \left(1 + \sum_{i=0}^{N-1} (i+1)[b_{j+i,j+i+1} + b_{j+i,j+i+2}] \right) + C_{1},$$

$$y_{j}^{(2)} = u_{j}b_{j,j+2} + \frac{A}{N} \left[\sum_{i=0}^{N-1} (i+1)b_{j+i+1,j+i+2} + b_{j+i+1,j+i+3} \right] + C_{2},$$
(32)

where the coefficients C_1 and C_2 are arbitrary constants that cancel in the final expression for dispersion. Then the final expressions for $T_{j,k}$ are found to be [23, 24]

$$T_{j,j+1} \left[1 - \left(\prod_{j=0}^{N-1} \frac{w_j}{u_j}\right)^2 \right] = \frac{-1}{u_{j+1}} \left[y_{j+1}^{(1)} + \sum_{k=1}^{N-1} y_{j+k+1}^{(1)} \prod_{i=j}^{j+k-1} \left(\frac{w_{i+1}w_{i+2}}{u_iu_{i+2}}\right) + \frac{w_{j+2}}{u_j} \left(y_j^{(2)} + \sum_{k=1}^{N-1} y_{j+k}^{(2)} \prod_{i=j}^{j+k-1} \left(\frac{w_{i+1}w_{i+3}}{u_{i+1}u_{i+2}}\right) \right) \right];$$

$$T_{j,j+2} \left[1 - \left(\prod_{j=0}^{N-1} \frac{w_j}{u_j}\right)^2 \right] = \frac{-1}{u_j} \left[y_j^{(2)} + \sum_{k=1}^{N-1} y_{j+k}^{(2)} \prod_{i=j}^{j+k-1} \left(\frac{w_{i+1}w_{i+3}}{u_{i+1}u_{i+2}}\right) + \frac{w_{j+1}}{u_{j+2}} \left(y_{j+2}^{(1)} + \sum_{k=1}^{N-1} y_{j+k+2}^{(1)} \prod_{i=j}^{j+k-1} \left(\frac{w_{i+2}w_{i+3}}{u_{i+1}u_{i+3}}\right) \right) \right].$$
(33)

For simplicity the trailing subunit is chosen as a marker for derivation of the explicit expressions for the drift velocity and dispersion. It can be shown that the same results are obtained if the centre of mass is used. The position of this particle at any time is given by

$$\langle x(t) \rangle = \frac{d}{N} \sum_{l=-\infty}^{\infty} \sum_{j=0}^{N-1} (j+Nl) [P_{j,j+1}(l,t) + P_{j,j+2}(l,t)]$$
$$= \frac{d}{N} \sum_{j=0}^{N-1} [C_{j,j+1}(t) + C_{j,j+2}(t)].$$
(34)

Utilizing this result along with the Master equations (11) we get

$$\frac{\mathrm{d}\langle x(t)\rangle}{\mathrm{d}t} = \frac{d}{N} \sum_{l=-\infty}^{\infty} \sum_{j=0}^{N-1} (j+Nl) [u_{j-1}P_{j-1,j+1}(l,t) - u_jP_{j,j+2}(l,t)].$$
(35)

Then the average drift velocity (2) has a simple form $V = \frac{d}{N}A$ where the function A is given by equation (29). The final expression for the velocity is

$$V = d \frac{\left[1 - (\prod_{j=0}^{N-1} \frac{w_j}{u_j})^2\right]}{R_N}.$$
(36)

A similar approach can be used to determine the dispersion, which can be done with the help of the following equation:

$$\langle x^{2}(t) \rangle = \frac{d^{2}}{N^{2}} \sum_{l=-\infty}^{+\infty} \sum_{j=0}^{N-1} (j+Nl)^{2} [P_{j,j+1}(l,t) + P_{j,j+2}(l,t)].$$
(37)

The time evolution of this quantity, again applying the Master equations (11), is given by

$$\frac{\mathrm{d}\langle x^2(t)\rangle}{\mathrm{d}t} = 2\left(\frac{d}{N}\right)^2 \sum_{j=0}^{N-1} \left[u_j C_{j,j+2}(t) - w_j C_{j,j+1} + \frac{1}{2} (u_j B_{j,j+2} + w_j B_{j,j+1}) \right].$$
(38)

Then after some algebra the expression for dispersion is written as

$$D = \left(\frac{d}{N}\right)^2 \sum_{j=0}^{N-1} \left[u_j T_{j,j+2} - w_j T_{j,j+1} + \frac{1}{2} (u_j b_{j,j+2} + w_j b_{j,j+1}) - A(T_{j,j+1} + T_{j,j+2}) \right].$$
(39)

And using the definition (3) we obtain the final formula:

$$D = \frac{d}{N} \left(\frac{dU_N + VS_N}{R_N^2} - \frac{N+2}{2} V \right),$$
 (40)

where

$$U_{N} = \sum_{j=0}^{N-1} \left[s_{2}(j)u_{j}r_{j,j+2} + s_{1}(j)\frac{A}{N}(R_{N} - Nr_{j-1,j+1}) \right],$$

$$S_{N} = \sum_{j=0}^{N-1} (s_{1}(j) + s_{2}(j-1))\sum_{i=0}^{N-1} (i+1)[r_{j+i,j+i+1} + r_{j+i,j+i+2}],$$

$$s_{1}(j) = \frac{1}{u_{j}} \left[1 + \sum_{k=1}^{N-1} \prod_{i=j}^{j-k+1} \frac{w_{i-1}w_{i}}{u_{i-2}u_{i-1}} + \frac{w_{j-1}}{u_{j-2}} \left(1 + \sum_{k=1}^{N-1} \prod_{i=j}^{j-k+1} \frac{w_{i-2}w_{i}}{u_{i-3}u_{i-1}} \right) \right],$$

$$s_{2}(j) = \frac{1}{u_{j}} \left[1 + \sum_{k=1}^{N-1} \prod_{i=j}^{j-k+1} \frac{w_{i}w_{i+2}}{u_{i-1}u_{i+1}} + \frac{w_{j+2}}{u_{j+1}} \left(1 + \sum_{k=1}^{N-1} \prod_{i=j}^{j-k+1} \frac{w_{i}w_{i+1}}{u_{i-1}u_{i}} \right) \right].$$
(41)

In this mechanism the bounds on the randomness parameter r can be estimated by calculating the dynamic properties for the simple limiting case when $u_j = u$ and $w_j = 0$ for all j. Then equations (25), (26) and (36) yield

$$R_N = \frac{2N}{u}, \qquad V_{\rm inch} = d\frac{u}{2N}, \tag{42}$$

while for dispersion we get from equations (40) and (41)

$$S_N = \frac{2N^2(N+1)}{u^2}, \qquad U_N = \frac{3N}{2u}, \qquad D_{\rm inch} = \left(\frac{d}{N}\right)^2 \frac{u}{8}.$$
 (43)

This analysis leads to r = 1/2N, which is the smallest possible value for this parameter. For any other set of transition rates $\{u_j, w_j\}$ the velocity is always smaller and the dispersion is larger, giving the general inequality for the inchworm mechanism

$$r_{\rm inch} \geqslant \frac{1}{2N}.$$
 (44)

Although we considered here only the case of m = 2, the method can be extended to include the inchworm models where the particles can be found more than two states apart.

2.4. Comparison of two mechanisms

The existence of exact analytical expressions for the dynamic properties of motor proteins with two heads in the hand-over-hand and the inchworm mechanisms allows us to analyse and compare these mechanisms very efficiently.

Consider first the simplest N = 1 models. Then the average velocity and dispersion for the hand-over-hand mechanism is given by

$$V_{\rm hoh} = d(u - w), \qquad D_{\rm hoh} = d^2(u + w)/2, \qquad (45)$$

The corresponding expressions for the inchworm mechanism can be obtained from equations (25), (26), (36), (40) and (41)

$$V_{\rm inch} = d(u - w)/2,$$
 $D_{\rm inch} = d^2(u + w)/8.$ (46)



Figure 4. Force–velocity curves for N = 2 model with $u_0 = 10 \text{ s}^{-1}$, $u_1 = 100 \text{ s}^{-1}$, $w_0 = 1 \text{ s}^{-1}$, $w_1 = 10 \text{ s}^{-1}$, $\theta_0^+ = \theta_0^- = \theta_1^+ = \theta_1^- = 0.25$ and d = 8.2 nm. The solid line describes the hand-over-hand mechanism, while the dashed line corresponds to the inchworm mechanism.

Thus the mean velocity in the inchworm model is only half of the velocity in the hand-over-hand mechanism, while the inchworm dispersion is only a quarter of the hand-over-hand value.

A more interesting case is N = 2 models where the average velocity and dispersion for the hand-over-hand mechanism are [32, 20, 21]

$$V_{\text{hoh}} = d(u_0 u_1 - w_0 w_1) / \sigma, \qquad D_{\text{hoh}} = \frac{1}{2} d^2 [(u_0 u_1 + w_0 w_1) - 2(V_{\text{hoh}}/d)^2] / \sigma, \qquad (47)$$

where $\sigma = u_0 + u_1 + w_0 + w_1$. For the inchworm mechanism the expression for the mean velocity can be written as

$$V_{\rm inch} = d \frac{(u_0 u_1)^2 - (w_0 w_1)^2}{2\sigma (u_0 u_1 + w_0 w_1) + (u_0 w_0 - u_1 w_1)(u_0 + w_0 - u_1 - w_1)},$$
(48)

while the explicit formula for dispersion is very bulky and it will not be presented here. Instead, we analyse the dependence of the dynamic properties of motor proteins on external forces using equations (5).

For illustration, the force–velocity curves for different mechanisms are presented in figure 4. It can be seen that the velocity for the inchworm mechanism is always smaller then the corresponding curve for the hand-over-hand mechanism, although the stall forces are the same. This can be explained by recalling that the stall force is a thermodynamic parameter for the sequential chemical kinetic models [21]. It is equal to the free energy difference between two consecutive binding sites divided by the step size d. Both the free energy difference and the step size are the same for the hand-over-hand and the inchworm mechanism, and this leads to the same value of the stall force.

The properties of dispersions for two mechanisms at different external loads, as shown in figure 5, are similar to the mean velocities. The particles that move via the inchworm mechanism fluctuate much less than the motor proteins utilizing the hand-over-hand method. This behaviour is expected since one of the motor subunits lowers the stochastic fluctuations of another motor head in the inchworm mechanism.

It is also interesting to compare the dimensionless function randomness for each mechanism: see figure 6. These results suggest that the motor proteins in the inchworm mechanism move more slowly and fluctuate less than the particles in the hand-over-hand mechanism, but the relative decrease in the fluctuations is larger than the relative lowering of



Figure 5. Dispersion as a function of external loads for different mechanisms of the motion. The solid line describes the hand-over-hand mechanism, while the dashed line corresponds to the inchworm mechanism. The parameters are the same as in figure 4.



Figure 6. Randomness at different external forces. The solid line describes the hand-over-hand mechanism, while the dashed line corresponds to the inchworm mechanism. The parameters are the same as in figure 4.

the average speed. This observation, that is correct for *any* N, is very important and it can be used for the experimental discrimination between different mechanisms of motor protein motility.

We propose the following procedure to determine the mechanism of the motion of motor proteins using *only* experimental measurements. First, from the independent bulk biochemical kinetic experiments determine the number of rate-limiting intermediate states. This information provides the size of the period, i.e., the parameter N. Second, from the high-precision single-molecule trajectories extract the velocity and dispersion for different (ATP) and different external forces. Such data can be obtained from the single-molecule optical trap experiments [6–11]. In the final step, analyse the randomness. If for some system this procedure yields r < 1/N, and the known number of intermediate states is N, it indicates that the motor protein cannot move by the hand-over-hand mechanism. However, for r > 1/N both mechanisms are still possible.

3. Summary and conclusions

The dynamics of motor proteins that move along the linear molecular tracks is discussed by taking into account the molecular structure and analysing in detail two possible mechanisms of motility. The motor proteins are viewed as two interacting particles that correspond to different motor domains in many conventional molecular motors [3]. The explicit expressions for the velocity and dispersion are obtained for the hand-over-hand mechanism, when the motor heads pass each other in the alternate fashion, and for the inchworm mechanism, when one motor domain is always ahead of the other one.

The exact calculation of the dynamic properties of molecular motors in the hand-overhand mechanism is performed by mapping the dynamics of two particles into the one-particle system, for which the dynamic properties are known exactly. It shows that in this case the dynamics is identical to the motion of the single free motor domain on the same biochemical pathway. The situation is very different for the inchworm mechanism. In this case we derived the exact analytic expressions for the velocity and dispersion by generalizing the single-particle Derrida's method [32, 21, 23, 24] to the system with two interacting particles.

Comparing the dynamics of molecular motors in two different modes, we conclude that the proteins in the inchworm mechanism move more slowly and fluctuate less than the particles in the hand-over-hand mechanism. Our results also indicate that the relative decrease in dispersion, expressed via the randomness parameter, is smaller for the inchworm mechanism. We suggest using this observation for the analysis of experimental data on motor proteins. The method of possible discrimination between two mechanisms of motor protein motility based on experimental observations is presented. In addition, the effect of external forces on the dynamic properties of molecular motors in the two mechanisms is also discussed.

The dynamic properties of motor proteins that move through the inchworm mechanism are obtained via the two-particle calculations. However, the average velocity and dispersion could also be obtained by mapping the system with two motor domains into the system with only one particle, for example, the centre of mass of the molecule. In general, the inchworm model where the distance between the individual motor domains is not larger than m sites can be mapped into the motion of a single particle on m - 1 parallel biochemical pathways, for which the dynamic properties are known exactly [25].

Our analysis of motor protein dynamics is rather very simplified since we considered molecules in which subunits interact only through the hard-core exclusion potential. However, the heads in motor proteins coordinate their motion and thus interact much more strongly than might otherwise be expected [3, 31]. It will be interesting to investigate the motor proteins with more realistic interactions between the subunits. The theoretical method used here seems capable of investigating more realistic systems of molecular motors.

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References

- Lodish H A, Berk A, Zipursky S L and Matsudaira P 1995 Molecular Cell Biology 3rd edn (New York: Scientific American Books)
- [2] Bray D 2001 Cell Movements: From Molecules to Motility 2nd edn (New York: Garland Publishing)

- [3] Howard J 2001 Mechanics of Motor Proteins and the Cytoskeleton (Sunderland, MA: Sinauer Associates)
- [4] Svoboda K, Mitra P P and Block S M 1994 Proc. Natl Acad. Sci. USA 91 11782
- [5] Kozielski F et al 1997 Cell **91** 985
- [6] Schnitzer M J and Block S M 1997 Nature 388 386
- [7] Kojima H, Muto E, Higuchi H and Yanagida T 1997 Biophys. J. 73 2012
- [8] Visscher K, Schnitzer M J and Block S M 1999 Nature 400 184
- [9] Mehta A D, Rock R S, Rief M, Spudich J A, Mooseker M S and Cheney R E 1999 Nature 400 590
- [10] Schnitzer M J, Visscher K and Block S M 2000 Nat. Cell. Biol. 2 718
- [11] Nishiyama M, Muto E, Inoue Y, Yanagida T and Higushi H 2001 Nat. Cell Biol. 3 425
- [12] Asbury C L, Fehr A N and Block S M 2003 Science 302 2130
- [13] Yildiz A, Tomishige M, Vale R D and Selvin P R 2003 Science 302 676
- [14] Mallik R, Carter B C, Lex S A, King S J and Gross S P 2004 Nature 427 649
- [15] Forkey J N, Quinlan M E, Shaw M A, Corrie J E T and Goldman Y E 2003 Nature 422 399
- [16] Snyder G E, Sakamoto T, Hammer J A, Sellers J R and Selvin P R 2004 Biophys. J. 87 1776
- [17] Oiwa K and Sakakibara H 2005 Curr. Opin. Cell Biol. 19 98
- [18] Qian H 1997 Biophys. Chem. 67 263
- [19] Jülicher F, Ajdari A and Prost J 1997 Rev. Mod. Phys. 69 1269
- [20] Kolomeisky A B and Widom B 1998 J. Stat. Phys. 93 633
- [21] Fisher M E and Kolomeisky A B 1999 Proc. Natl Acad. Sci. USA 96 6597
- [22] Fisher M E and Kolomeisky A B 1999 Physica A 274 241
- [23] Kolomeisky A B and Fisher M E 2000 Physica A 279 1
- [24] Kolomeisky A B and Fisher M E 2000 J. Chem. Phys. 113 10867
- [25] Kolomeisky A B 2001 J. Chem. Phys. 115 7253
- [26] Fisher M E and Kolomeisky A B 2001 Proc. Natl Acad. Sci. USA 98 7748
- [27] Mogilner A, Fisher A J and Baskin R J 2001 J. Theor. Biol. 211 143
- [28] Bustamante C, Keller D and Oster G 2001 Acc. Chem. Res. 34 412
- [29] Kolomeisky A B and Fisher M E 2003 Biophys. J. 84 1642
- [30] Lan G and Sun S X 2005 *Biophys. J.* 88 999
- [31] Stukalin E B, Phillips H and Kolomeisky A B 2005 Phys. Rev. Lett. 94 238101
- [32] Derrida B 1983 J. Stat. Phys. 31 433